Three new marine ascomycetes from driftwood in Australian sand dunes

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Caryospora australiensis sp. nov., *Platystomum scabridisporum* sp. nov. and *Savoryella melanospora* sp. nov. are described from driftwood collected from a sand dune at Rye, on the Mornington Peninsula Nature Park, Victoria, Australia. These species are compared with other taxa in these and related genera.

Key Words—arenicolous; marine fungi; taxonomy.

Early studies on the marine fungi of Australia were carried out by Cribb and Cribb (1955, 1956, 1960, 1969). More recently Hyde (1991, 1992, 1993a, b) and Koh-Imeyer and Volkmann-Kohlmeyer (1991a) have documented the occurrence of marine fungi from the tropical waters of Queensland to the subtropical shores in the Melbourne area. Hyde (1996) reviewed our knowledge of marine fungi from Australia, listing 100 species, extensively illustrating the fungi found and with suggestions for future research. The Mornington Peninsula Nature Park has a well-developed sand dune system with a significant amount of driftwood buried in the sand. Wood was collected from such a habitat and examined for the presence of marine fungi.

Material and Methods

Wood buried in sand was collected in the Mornington Peninsula Nature Park, Rye, Victoria, Australia. Material collected was returned to the laboratory in Hong Kong where it was incubated in sterile, humid chambers at room temperature. Examination of material follows the well-adopted procedures described by Jones and Hyde (1988).

Taxonomy

Caryospora australiensis Abdel-Wahab & E. B. G. Jones, sp. nov. Figs. 1-9

Etym.: *australiensis* in reference to the place where the fungus was first collected.

Ascomata 870–1000 μ m alta, 830–850 μ m diam, late, conica vel subglobosa, basaliter complanata vel rotundata, immersa vel erumpentia, carbonacea, papillata, centro ostiolata, solitaria vel gregaria. Peridium bistratum; stratum exterius ad apicem 110–190 μ m crassum et basi 72–177 μ m crassum, brunneum vel nigrum. Trabeculae ad apicem 1.2–2 μ m diam, supra ascos anastomosantes, periphysibus transientes, in matrice gelatinosa inclusae. Asci 162–215 × 20–30 μ m, cylindrici vel subcylindrici, fissitunicati, pedunculati, octospori. Ascosporae 30–44 × 12–17 μ m, uni-vel biseriatae, triseptatae, nigrae, constrictae vel non constrictae, ellipsoideae, striatae.

Holotypus: In ligno indet., Mornington Peninsula, Rye, Victoria, Australia, Aug. 1998, coll. E. B. G. Jones, IMI 379748.

Ascomata 870-1000 µm high, 830-850 µm in diam, broadly conical or subglobose, with a flat or rounded base, immersed to erumpent, carbonaceous, papillate, with a central ostiole, solitary or gregarious (Fig. 1). Papillae 85-200 µm long, 100-155 µm in diam, ostiolar canal 60–180 μ m long and 50–100 μ m in diam, periphysate; periphyses $1.5 \,\mu m$ in diam (Figs. 1, 3). Peridium two-layered: outer layer carbonaceous, in the superficial part of the ascoma 110-190 μ m thick, composed of heavily melanized angular pseudoparenchymatous cells; in the emmersed part of the ascoma 72-177 μ m thick, composed of brown to dark brown angular or horizontally oriented palisade-like cells; host cells occasionally incorporated in large amount inside the peridial wall; cells in the basal area acute angle of the cone palisade-like, vertically oriented, brown to dark brown in colour; inner layer 12-20 μ m thick throughout the ascomatal cavity and composed of polygonal hyaline cells (Fig. 4). Trabeculae 1.2-2 μ m in diam at the tip, anastomosing above the asci, merging with the periphyses in the ostiolar canal, and embedded in a gelatinous matrix (Figs. 2, 6). Asci $162-215 \times 20-30 \ \mu m$ ($\bar{X}=181.8 \times 23.7 \ \mu m$, n=30), peripheral, cylindrical to subcylindrical, fissitunicate, pedunculate, eight-spored, and with a wide ocular chamber (Figs. 5, 6). Ascospores 30–44 \times 12–17 μ m



Figs. 1-4. *Caryospora australiensis*, differential interference light micrographs.
1. Longitudinal section through aggregated ascomata, each one with a papilla and ostiole.
2. Magnified part of a longitudinal section through an ascoma showing trabeculae anastomosing above the asci (arrowed) and embedded in a gelatinous matrix.
3. Longitudinal section through the ostiolar canal, showing the periphyses (arrowed) filling the ostiolar canal cavity.
4. Section through the peridium with melanized black outer stratum (arrowed) and inner stratum consisting of polygonal hyaline cells. Bar lines: Fig. 1=240 μm; Figs. 3, 4=20 μm.

 $(\bar{X}=38.2 \times 14.9 \ \mu m, n=50)$, uni- or biseriate, 3-thickened eu-septate, the end cells faint in colour, all the cells becoming black when mature, constricted or not at the central septum, ellipsoid, and smooth-walled, the median cells becoming striate with age (Figs. 7-9).

Type material: IMI 379748 Aug. 1998, on driftwood buried in sand, at the base of sand dune, at Mornington Peninsula, Rye, Victoria, Australia. coll. E. B. G. Jones

Table 1.	Morphological	comparison of	Caryospora	australiensis,	С.	langloisii and	С.	striata
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	C. australiensis	C. langloisii (Barr. 1979)	<i>C. striata</i> (Barr 1979)
Ascomata	870–1000 μm high, 830–850 μm in diam, black	1 mm in diam, dark brown	660 μ m in diam, dark brown
Peridium	72–177 μ m thick at the base, 112– 192 μ m thick at the sides and the upper half of the ascomata	ca. 50 μm wide at base, up to 78 μm wide at sides	ca. 30 μm wide, thickened toward exposed apex
Asci	162–215×20–30 μm, cylindrical to subcylindrical, with wide ocular chamber	110–170×40–45 μ m, broadly oblong	240×18−22 μm
Ascospores	30–44 × 12–17 μ m (\bar{X} =38.2 × 14.9 μ m), 3-septate becoming opaque in age, constricted or not at the central septum, ellipsoid, smooth-walled becoming striate at maturity, especially the mid-cells	32–50 × 15–20 μ m (\bar{X} = 41 × 17.5 μ m), rich reddish brown, ellip- soid fuscoid, primary septum median, slightly constricted, secondary septa near each tip, not constricted, with one large globule in each cell, sur- rounded by narrow gelatinous sheath, smooth, some showing wrin- kling of surface especially at the apex	$40-50 \times 14-15 \ \mu m$ ($\bar{X} = 45 \times 14.5 \ \mu m$), dark reddish brown, ellipsoid, primary septum median, constricted, no additional septa seen, contents with one large and one small globule in each cell, wall finely roughened





Figs. 5–9. Caryospora australiensis, differential interference light micrographs.
5, 6. Immature (Fig. 5) and mature (Fig. 6) asci, note the ocular chamber in the mature ascus. 7–9. Ascospore 3-septate, fuscous, and intensively black at maturity. Bar lines: Figs. 5, 6=15 μm; Figs. 7–9=5 μm.



Figs. 10-13. Platystomum scabridisporum, differential interference light micrographs.

Longitudinal section through a mature ascoma embedded in a woody substratum. 11. Magnified part of centrum with pseudoparaphyses, branched and anastomsing above the asci (arrowed), merging with the periphyses in the ostiolar canal. 12, 13. Section through the peridial wall with an outer stratum (arrowed OS) of thick walled melanized cells, interspersed with cells of the host; and the inner stratum (arrowed IS) of hyaline rounded cells. Bar lines: Figs. 10, 11=100 μm; Figs. 12, 13=15 μm.



Figs. 14-22. Platystomum scabridisporum, differential interference light micrographs.

14, 15. Immature (Fig. 14) and mature (Fig. 15) asci, note apical ocular chamber.
 16. Fissitunicate dehiscence of the ascus.
 17. Hyaline trabeculae.
 18–22. Morphological variation of ascospores with verrucose spore wall (arrowed Fig. 18). Bar lines: Figs.
 14, 15, 17=3 μm; Fig. 16=25 μm; Figs. 18–22=5 μm.

(Holotype).

Habitat: Saprobic on driftwood. Distribution: Australia.

Of the six described Caryospora De Not. species, C. australiensis most closely resembles C. langloisii Ellis & Everh. and C. striata (Niessl) Scheinpflug. Caryospora australiensis differs from C. langloisii in having cylindrical, longer and narrower asci and smaller ascospores without a gelatinous sheath. Caryospora australiensis differs from C. striata in having larger ascomata and smaller ascospores, while it differs from both species in having 3-thickened eu-septate ascospores becoming black at maturity (Table 1) (Barr, 1990). Hyde (1989) described Caryospora mangrovei K. D. Hyde from mangrove wood collected in Brunei with 1-3-septate hazel brown ascospores measuring 36-60 \times 16-24 μ m, but Kohlmeyer and Volkmann-Kohlmeyer (1991b) transferred this species to Coronopapilla mangrovei (K.D. Hyde) Kohlm. & Volkm.-Kohlm.

Platystomum scabridisporum Abdel-Wahab & E.B.G. Jones, sp. nov. Figs. 10-22

Etym.: From the Latin *scabridus* meaning "with delicate projections" in reference to the verrucose ascospore cell wall and *-sporum* meaning "-spored".

Ascomata 465–740 μ m alta, 400–650 μ m lata, subglobosa, immersa, erumpentia, ostiolata, nigra, coriacea, solitaria vel gregaria. Peridium bistratum; stratum exterius 20-42 μ m crassum, nigrum, textura angulari formatum, cellulis hospitis includens; stratum intimium 15-32 µm crassis, ex cellulis parvis hyalinis pseudoparaphysibus transientibus compositum; ostiolum depressum in centro sulci apicale, circulare; canalis ostioli 220-350 μ m longus et 60–100 μ m diam, cylindricus, reticulo periphysium ramosarum septatarum usque 60 µm longarum et 1-2 μ m diam impletus. Trabeculae 1.2-2 μ m diam, ad basim eramosae, supra ascos ramosae et anastomosantes, periphysibus transientibus in matrice gelatinosa inclusae. Asci 105–170 \times 12–15 μ m, cylindrici, pedunculati, octospori, fissitunicati. Ascosporae 20 -32×7 -11 μ m, fusiformes, muriformes, 5-7(-8) transseptatae, 1-2(-3) longiseptatae, brunneae, scabridae.

Holotypus: In ligno indet., Mornington Peninsula, Rye, Victoria, Australia, Aug. 1998, coll. E. B. G. Jones, IMI 379750.

Ascomata 465-740 μ m high, 400-650 μ m in diam $(\bar{X} = 569.5 \times 464 \,\mu m, n = 10)$, subglobose, immersed, papillate, erumpent, ostiolate, black, coriaceous, and solitary or gregarious (Fig. 10); Papillae beaked, 120-235 μ m wide, 220–350 μ m long. Peridium two-layered (Figs. 12, 13): outer layer 20-42 μ m thick at the base and the sides, 60-135 μ m thick near the ostiole, composed of melanized cells forming textura angularis, interspersed with cells of the host; and the innermost laver 15–32 μ m thick at the base, 5–10 μ m thick at the sides which is absent in the upper part of ascomatal cavity and ostiolar canal, composed of hyaline rounded or weakly angular small cells that merge with the pseudoparaphyses. Ostiole depressed in the center of the apical furrow, slit-like with an ostiolar canal 220–350 μ m long, 60 $-100 \,\mu\text{m}$ wide, cylindrical, filled with a network of branched, septate periphyses up to 60 μ m long, 1–2 μ m in diam at their tips (Figs. 10, 11). Trabeculae 1.2–2 μ m in diam, unbranched at the base, becoming branched and anastomosing above the asci, merging with the periphyses in the ostiolar canal, and embedded in a gelatinous matrix (Figs. 11, 17). Asci 105-170×12-15 μ m (\bar{X} = 136.2 × 13 μ m, n = 30), cylindrical, pedunculate, eight-spored, fissitunicate, with an apical ocular chamber (Figs. 14–16). Ascospores $20-32 \times 7-11 \, \mu m$ $(X = 25.3 \times 8.7 \,\mu m, n = 50)$, fusiform, uni- or biseriate, muriform, with 5-7(-8) transverse septa and 1-2(-3) longitudinal septa, constricted at the central septum, brown with a rough or verrucose spore wall surface (Figs. 18-22).

Type material: IMI 379750, Aug. 1998, on driftwood buried in sand, at the base of a sand dune, at Mornington Peninsula, Rye, Victoria, Australia, coll. E. B. G. Jones (Holotype).

Habitat: Saprobic on driftwood. Distribution: Australia. Our collection is assigned to the genus *Platystomum*

-	P. scabridisporum	P. compressum (Barr, 1990)	P. obtectum (Barr, 1990)
Ascomata	465–740 μm high, 400–650 μm in diam, black	440–725 μ m in diam	440-770(-1000) μm in diam
Papilla	Peaked, 120-235 µm wide, 220 -350 µm long	220 μm wide and high, com- pressed, surrounded by brown hyphae	Compressed or occasionally rounded or triangular, 110– 165 μm wide and high
Peridium	35–74 μ m, up to 135 μ m wide	30–50 μ m wide	25–50 μ m, up to 90 μ m wide
Asci	Cylindrical, 105–170×12– 15 µm, 8-spored	Cγlindrical, 100–160×10– 15 μm, 8-spored	Clavate, (80–)100–140×12– 20 μm, 4-8-spored
Ascospores	$20-32 \times 7-11 \ \mu m \ (\overline{X} = 25.3 \times 8.7 \ \mu m)$, brown, 5-7(-8) transverse septa, 1-2(-3) longitudinal septa, rough spore wall surface, without gelatinous sheath	$(15-)18-28\times 6-8-(-9) \ \mu m$ $(X=23\times 7 \ \mu m)$, reddish brown, $(3-4-)5-(6-7-8)$ septa, one longitudinal septum in the mid cells, wall smooth, surrounded by a gelatinous sheath	$(18-)20-32 \times (6-)7-10-(-11)$ μ m (X=26 × 9.3 μ m), clear brown to reddish brown, (3-)5 (-7) septa, one longitudinal sep- tum in one or a few mid cells, wall smooth or verrculose, sur- rounded by gelatinous sheath

Table 2. Morphological comparison of Platystomum scabridisporum, P. compressum and P. obtectum.



Figs. 23-26. Savoryella melanospora, differential interference light micrographs.

23. Longitudinal section through an ascoma. 24. Peridial wall with an outer stratum of melanized thick-walled, dark brown to black cells; and an inner stratum of hyaline to yellow brown, elongated cells. 25. Longitudinal section through the neck with periphyses. 26. Paraphyses. Bar lines: Fig. $1=50 \ \mu\text{m}$; Figs. $2-4=15 \ \mu\text{m}$.



Figs. 27–33. Savoryella melanospora, differential interference light micrographs.
 27–29. Immature and mature asci with apical thickening (arrowed in Fig. 29). 30–33. Dark brown to fuscous ascospores (central cells) with hyaline to sub-hyaline end cells. Bar lines: Figs. 27, 28=20 μm; Figs. 29–33=5 μm.

Trev., Platystomaceae, Melanommatales (Barr, 1990) in the P. compressum (Pers.: Fr.) Trev. group. Platystomaceae is based on the genus Platystomum Trev. The type species of this genus, P. compressum, however has usually been treated as a Lophiostoma species (Holm and Holm, 1988). Eriksson and Hawksworth (1991) suggested that *Platystomum* is kept as a synonym of Lophiostoma, and in accordance with Holm and Holm (1988) the Platystomaceae is treated as a synonym of Lophiostomataceae. However, we assign our species to Platystomum as the ascomata are immersed, becoming erumpent with apex well developed, papillate, pore slit-like, asci cylindrical and narrow, trabeculae in a gel matrix and with brown, muriform ascospores. The fungus resembles the marine bitunicate genus Aigialus Kohlm. & S. Schatz but differs in the smaller, narrower asci without the well-developed ascus apparatus and fewer septate ascospores which are not flattened and lack a gelatinous cap (Kohlmeyer and Schatz, 1985).

The genus *Platystomum* comprises 6 species (Chester and Bell, 1970; Barr, 1990). Our species most closely resembles *P. compressum* and *P. obtectum* Peck but differs in the smaller ascomata, longer neck, wider peridium, longer asci and in the greater number of transverse and longitudinal septa to the ascospores (Table 2). In *P. compressum* and *P. obtectum* the ascospores are surrounded by a mucilaginous sheath which is absent in *P. scabridisporum*. The verrucose wall of *P. scabridisporum* is clearly visible in Figs. 18 and 19.

Savoryella melanospora Abdel-Wahab & E. B. G. Jones, sp. nov. Figs. 23-33

Etym.: From the Greek *melan* meaning "black" in reference to the intensively black ascospores and -spora meaning "-spored".

Ascomata 160–305 μ m alta, 184–225 μ m lata, globosa vel subglobosa, obliqua vel horizontalia, solitaria vel gregaria, semi-immersa vel superficialia, ostiolata, papillata. Colla 80–275 × 70–160 μ m, lateralia, periphysata. Peridium bistratum, textura angulari formans. Asci 170–212 × 15–25 μ m, octospori, cylindrici, pedunculati, unitunicati, persistentes, ad apicem truncati et annulo crasso in amyloideo praediti. Ascosporae 32– 45 × 15–18 μ m, ellipsoideae, uniseriatae, triseptatae, cellulis centralibus nigris, cellulis polaribus subhyalinae.

Holotypus: In ligno indet., Mornington Peninsula,

Rye, Victoria, Australia, Aug. 1998, coll. E. B. G. Jones, IMI 379749.

Ascomata 160–305 μ m high, 184–225 μ m in diam, globose to subglobose, oblique to horizontal, solitary or gregarious, partly immersed to superficial, ostiolate, papillate, membranous and dark brown to black (Fig. 23). Neck 80-275 μ m long, 70-160 μ m in diam, lateral, brown, lighter and slightly tapering towards the apex (Fig. 25). Ostiolar canal 50–80 μ m in diam, and periphysate; periphyses 20-25 μ m long, 0.5 μ m in diam (Fig. 25). Peridium two-layered, forming textura angularis (Fig. 24); outer layer 7-17 μ m thick, 2-4 layers of melanized thick-walled, dark brown to black cells and with brown septate hyphae around the ascomata and necks, 2–2.5 μ m in diam; inner layer 10–20 μ m thick, 7– 9 layers of hyaline to yellow brown, elongated cells which merge with the paraphyses. Paraphyses 5–19 μ m wide, hyaline, septate and deliguescing early (Fig. 26). $170-212 \times 15-25 \ \mu m$ $(\bar{X} = 190.5 \times 19.7 \ \mu m)$ Asci n=30), eight-spored, cylindrical, pedunculate, unitunicate, persistent, truncate at the apex, with a non-amyloid apical thickening, with a limited number of asci in each ascoma at different developmental stages (Figs. 27-29). Ascospores $32-45 \times 15-18 \ \mu m$ $(\bar{X} = 37.4 \times 16 \,\mu m)$ n=50), ellipsoid, uniseriate, 3-septate, slightly constricted at the septa, central cells black, end cells smaller and hyaline to subhyaline (Figs. 30-33).

Type material: IMI 379749, Aug. 1998, on driftwood partially buried in sand, at the base of a sand dune, at Mornington Peninsula, Rye, Victoria, Australia, coll. E. B. G. Jones (Holotype).

Habitat: Saprobic on driftwood.

Distribution: Australia.

Ten Savoryella E. B. G. Jones & R. A. Eaton species have been described (Ho et al., 1997; Chang et al., 1998), of which, S. lignicola E. B. G. Jones & R. A. Eaton, S. paucispora (Cribb & J. W. Cribb) J. Koch, S. longispora E. B. G. Jones & K. D. Hyde and S. appendiculata K. D. Hyde & E. B. G. Jones are marine. Savoryella melanospora most closely resembles S. aquatica K. D. Hyde that was described by Hyde (1993b) on submerged wood in a freshwater river in Australia. Savoryella melanospora differs from S. aquatica in its marine habitat and in having larger ascomata, asci and ascospores (Table 3). The ascospores of S. grandispora K. D. Hyde are larger (46–58 × 14–16 μ m) than those of S. melanospora,

Table 3. Morphological comparison of Savoryella aquatica and S. melanospora.

	S. melanospora	<i>S. aquatica</i> (Hyde, 1993b)
Ascomata	160–305 μm high, 184–225 μm in diam, black	195–260 μm high, 91–130 μm in diam, brown
Neck	50–80 μ m in diam	68 µm in diam
Asci	Cylindrical, 170–212×15– 25 µm	Cylindrical, 106−140×26− 34 µm
Ascospores	32–45 × 15–18 μ m (\bar{X} = 37.4 × 16 μ m), central cells fuscous, intensely black, end cells hyaline	29–38×13.5–17 μ m (\bar{X} =33.5 × 15.2 μ m), central cells brown, end cells hyaline

dark brown in colour and the former is a freshwater species (Hyde, 1993b).

Discussion

In this preliminary study of the marine fungi on driftwood collected in a well-developed sand dune system in the Mornington Peninsula National Park, Rye, Australia, three new species were collected. Other fungi collected were: Corollospora cinnamomea J. Koch (5 collections), a Diaporthe species, Leptosphaeria australiensis (Cribb & J. W. Cribb) G. C. Hughes, Aigialus sp., Linocarpon sp., Phoma sp. and a discomvcete with black apothecia. This suggests that we are dealing with marine sand dune fungi rather than a desert fungal community. Sand dunes are harsh habitats with exposed and buried wood subject to intense temperature variation, light intensity including UV, and subject to drying out at the collecting site. The sand dune vegetation was typical of the region and supports a number of low shrubs and herbaceous plants. Exposure to intense sunshine may account for the intense black colour (melanization) of the ascospores in the three species described in this paper, and the immersion of the bitunicate asci in C. australiensis and P. scabridisporum in a gelatinous matrix. The protection of asci in a gelatinous matrix is a feature of many bitunicate ascomycetes and discomycetes in mangrove habitats, where the ascomata are exposed for long periods, e.g. Dactylospora haliotrepha (Kohlm. & E. Kohlm.) Hafellner (Au et al., 1996; Kohlmeyer and Volkmann-Kohlmeyer, 1998), Melaspilea mangrovei Vrijmoed, K.D. Hyde & E. B. G. Jones (Vrijmoed et al., 1996) and many Massarina species (Read et al., 1994). The importance and role of this gelatinous matrix in the protection of developing sporulating structures warrants further investigation.

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